Package 'rcpphmmparclip'

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Type Package

Title Rcpp functions for analysing PAR-CLIP using the HMM

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Description This package provides essential functions for the forward-backward Gibbs sampler to snalyse PAR-CLIP data using the HMM.

Depends Rcpp (>= 0.9.3)

License GPL (>= 2.1)

R topics documented:

`rcpphmmparclip-package`
Rcpp functions for analysing PAR-CLIP using the HMM

Description

This package provides essential functions for the forward-backward Gibbs sampler to snalyse PAR-CLIP data using the HMM.

Details

To install this package, windows users may require to install Rtools (http://cran.r-project.org/bin/windows/Rtools/), which will help to compile the Rcpp functions in your machine. It is recommended to install both R and Rtools in directories whose names have no space. More details about how to build the package is provided in Rcpp-FAQ (http://dirk.eddelbuettel.com/code/rcpp/Rcpp-FAQ.pdf).

To install the package,

```
install.packages('rcpphmmparclip_0.1.0.tar.gz',type='source')
```

Author(s)

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References

Yun, J., Wang, T. and Xiao, G. (2013). Bayesian HMMs to Identify RNA-Protein Interaction Sites in PAR-CLIP. Biometrics (under revision).

See Also

bases, fbgibbs

Examples

```
require(rcpphmmparclip)
require(doMC)
require(parallel)

tcore = detectCores()
cl <- makeCluster( tcore )
registerDoMC()

data(bases)

mc = fbgibbs(bases,tot=100,nsv=tcore);

# print sites with the posterior probability >= postcut.
postcut = 0.8;
ranking(bases,mc,postcut);
# estimate FDR on these sites.
fdr(mc,postcut);

stopCluster(cl)
rm(cl)
```

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bases *An example PAR-CLIP data set*

Description

This is a short example of PAR-CLIP dataset. Aligned read seuqneces are converted to read and mutation counts in each genomic location. Only T to C substitutions are kept, and non-clustered reads are discarded.

Usage

```
data(bases)
```

Format

tag `integer` Read counts

mutant `integer` Mutation counts (T to C substitutions)

region_id `string` ID that is assigned to a CLIP cluster

chr `string` Chromosome

strand `string` Strand information

nt `string` Nucleotide sequence

pos `integer` Genomic location

Examples

```
require(rcpphmmparclip)
require(ggplot2)
data(bases)
attach(bases)
rid = 85974;
ridx = (region_id == rid);
rlen = sum(ridx);

x = rep(as.factor(pos[ridx]),2);
y = c(tag[ridx] - mutant[ridx],mutant[ridx]);
factor = as.factor(c(rep("read",rlen),rep("mutation",rlen)))
qplot(x,geom="bar", weight=y, fill = factor, xlab='Genomic Location',
      title='Graphical representation of a CLIP cluster')
```

`fbgibbs` *The forward-backward Gibbs sampler*

Description

This function estimates posterior probabilities of binding sites, posterior means of parameters. The function also provides the posterior predictive checking.

Usage

```
fbgibbs(data, c = 3, p = c(0.04, 0.02, 0.3), pi = c(0.5, 0.5, 0),
                     fkab = t(matrix(c(1,-1,0,
                                       1,1,0.05,
                                       1,1,0.05),3,3)),
                     K_T = matrix(c(0.98, 0.02, 0.00,
                                    0.05, 0.92, 0.03,
                                    0.00, 0.99, 0.01),3,3),
                     K_N2 = c(0.05, 0.95, 0.00),
                     phi_T = c(0.90, 0.1, 0.00),
                     epsil = 0.195, delta = 0.005, tot = 10000, burnp = 0.5,
                     nsv = 1, doppc = FALSE, doprintout = FALSE)
```

Arguments

data	PAR-CLIP data to be analysed. See ...
c	The truncation value for read counts.
p,pi	Initial values of mutation probabilities p_s and zero-inflated probabilities π_s.
fkab	Initial values of parameters in the beta geometric distribution.
K_T	Initial values of transition matirx K_T on T-sites.
K_N	Initial values of the 2nd row of the transition matrix K_N on non T-sites.
phi_T	Initial values of the initial distribution on T-sites of hidden Markov chains.
epsil	The upper bound ϵ of mutation probabilities on non-binding sites.
delta	The lower bound $\epsilon + \delta$ of mutation probabilities on binding sites.
tot	The maximum number of MCMC iterations.
burnp	The proportion of MCMC chains to be burnt out.
nsv	The number of cores to be used for parallel computing.
doppc	If TRUE, the posterior predictive checking is carried out. However, this option will slow down the FB Gibbs sampler.
doprintout	If TRUE, summeries for each MCMC chains will be provided.

Value

IP	[,3] Posterior probability of the binding site
ppc	The p-value for the posterior predtive checking
sumc	posterior means of parameters
sumc$bg	posterior means of parameters in the BG. [,2]: μ_s and [,3]: η_s

See Also

bases

Examples

```
require(rcpphmmparclip)
require(doMC)
require(parallel)

tcore = detectCores()
cl <- makeCluster( tcore )
registerDoMC()

data(bases)

mc = fbgibbs(bases,tot=100,nsv=tcore);

# print sites with the posterior probability >= postcut.
postcut = 0.8;
ranking(bases,mc,postcut);
# estimate FDR on these sites.
fdr(mc,postcut);

stopCluster(cl)
rm(cl)
```

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